



Effect of Cytokinin Type and pH Level on Regeneration of Ginger *in vitro*

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ABSTRACT

Ginger is unfertile species that usually failed to set seed. Cultivation of this plant using its economic part (rhizomes) is an unprofitable method, which negatively affects its supply in the market. Therefore, this study aimed to maintain the local Yemeni ginger variety, and facilitate its propagation by tissue culture technique, using fresh rhizome buds in a semi-rigid culture medium. To do so, two experiments were carried out, the first one was based on completely randomized block design with four replicates to determine the optimum pH levels (including 5.6, 5.8, 6.0) for the shoot and root formation. The result showed that the 5.8 is the optimum pH level of the medium. Second experiment was designed as two factors, type of cytokinin [6-benzylaminopurine (BAP) and Kiniten (Kin) (N⁶-furfuryladenine)] and ii) Cytokinin concentrations (0, 1, 2 and 3 mg L⁻¹). The MS basal medium supplemented with above types and concentration of cytokinin with 1.0 mg L⁻¹ IBA and 3% sucrose (BAP and Kiniten) The explants cultured on Murashige and Skoog's (MS) medium supplemented with cytokinin with four replicates. The result showed that the explants cultured on MS basal medium supplemented with 2.0 mg L⁻¹ Kin + 1.0 mg L⁻¹ IBA give the highest rate of shoot multiplication, shoot length and root number. In conclusion, the result obtained from this study might help to manage the propagation of ginger.

Introduction

Ginger (*Zingiber Officinale* Rosc.) is the most important spice plant in the Zingiberaceae family. Ginger production for the extraction of its active compound is oleoresins and essential oils, as well as the direct use of rhizomes for culinary purposes (Ayenew et al., 2012). In Yemen ginger is traditionally used as juice or as an additive to a coffee beverage. It is highly appreciated for its medicinal properties, such as anti-inflammatory, antifungal, analgesic and antioxidant content (Ozaki et al., 1991; Kishore and Dwivedi, 1992). Ginger rhizome uses post-natal treatment,

swelling, rheumatism, application of joint pain, intestinal disturbance, and numbness of the feet (Sirirugsa, 1999). Owing to the universal outbreak of COVID-19 virus, ginger consumption achieve more interest. Ginger helps to alleviate the severe symptoms of COVID-19 positive patients and reduce the recovery time in those patients (Zahid et al., 2021). The total world production and cultivated area of ginger have sharply increased from 1992 to 2020, where China and Thailand were the major exporting countries in 1998-2000. Furthermore, importing and exporting quantity (ton) in Yemen has stimulated as 6780 and 137 tons in 2013 FAOSTAT (2013).

Overall, the propagation of plants makes up the first stage of the horticultural economy by increas

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ng the cultivated area. Tissue culture techniques can be beneficial for quick clonal propagation and production of disease-free plants with completely similar to their mother plants (Eshghi et al., 2020). It has also been used effectively to protect important and endangered horticulture plants (Bhattacharyya and Kumaria 2014; Naderi et al. 2021).

Ginger does not produce seeds as it is an unfertilized species. It regularly propagates vegetatively via its rhizomes (Nair 2019). Using ginger rhizomes as a raw material for the next crop production is a non-economical method, and negatively affects its supply to the marketplace (Zahid et al., 2021). The suitable material for plantlet formation of ginger *in vitro* has been previously reported, which could be propagation by axillary buds, sprouting buds and shoot tips isolated from the rhizomes (Doraiswamy et al., 1983; Balachandran et al., 1990; Olivier, 1996).

The level of pH plays an important role in the manipulation or management of the explant induction outcome. Abdul Aziz et al. (2012) demonstrated that the optimal pH of the medium for sprout formation is 5.5-6.0. Meanwhile, regeneration of shoot buds requires the pH level of about 5.7 or 5.8 (Rout and Das, 1997).

Plant growth regulators are one of the most significant components of the growth and development medium in tissue culture techniques, wherein the cytokinin is the most important of these regulators because of its wide roles. The medium supplemented with different types and concentrations of the hormone for vegetative and rooting growth of ginger has yielded different results. Previous studies showed that the BAP and NAA combinations were the most effective for the growth of shoots in ginger (Sakamura et al., 1986; Charlwood et al., 1988; Sakamura and Suga, 1989).

Sharma and Singh (1997) reported that, the high-frequency *in vitro* multiplication of disease-free clones and successful transferring plantlets of ginger (*Zingiber Officinale* Rosc.) obtained by culturing small and active buds of ginger on MS medium supplemented with 2 mg L⁻¹ Kin and 20 g L⁻¹ sucrose. Nirmal (1997) tested various explants of ginger on MS basal medium supplemented with cytokinins (BAP and Kin) and auxins (NAA and 2, 4-D), the medium with 4 mg L⁻¹ NAA and 4 mg L⁻¹ BAP gave positive response in inducing multiple shoots and roots. Ayenew et al. (2012) found that 2 mg L⁻¹ BA + 1 mg L⁻¹ Kin was better than other combinations on shoot number of ginger, and NAA alone developed numerous longer roots.

Behera and Santilata (2009) reported that explants grown on a basal MS medium containing 2.0

mg L⁻¹ BAP + 0.5 mg L⁻¹ NAA had the highest rate of ginger stem propagation. Rooting of ginger *in vitro*, was better in the basal MS medium supplemented with 2.0 mg L⁻¹ NAA, and medium with 4 mg L⁻¹ BAP + 3 mg L⁻¹ Kin + 1 mg L⁻¹ IAA caused the best performance, whereas, MS medium with 2 mg L⁻¹ IBA + 2 mg L⁻¹ NAA was very effective on root initiation (Hossain et al., 2010). Eshghi et al. (2020) found that 5.4 μM NAA or 8.9 μM IBA is the most effective treatment for rooting of *Passiflora edulis* sprouts.

According to the aforementioned studies, the current investigation aimed to develop a reliable protocol for the *in vitro* propagation of the local Yemeni ginger variety in order to: i) reveal the best optimal pH levels predominantly used in the culture medium. ii) Determine the various levels of 6-benzylaminopurine (BAP) and Kinetin (N6-furfuryladenin) during the spread of the local Yemeni ginger variety. The findings could help to establish and spread cultivation of ginger in different environmental conditions in Yemen by providing healthy and economic seedling.

Materials and Methods

The present experiment was conducted in Plant Tissue Culture Unit, Faculty of Agriculture-Sana'a University, Yemen during the period from January 2012 to June 2012 and aimed to establish a suitable and reproducible protocol for *in vitro* regeneration of local ginger (*Zingiber Officinale* Rosco).

Plant materials and source collection

A local variety of Yemeni ginger applied as a plant substance in this experiment. It collected from Malhan highlands region located at west of Al-Mahwit province (15°28'10" N 43°32'43" E) north-west of capital city of Yemen (Fig. 1). Rhizomes preserved in a mixture of peat moss and sandy soil in the laboratory to get growing shoots used as explants. Rhizome buds 1-2 cm long selected as initial explants. Collected fresh buds cleaned and left under running water of the faucet for ½ - 1 h. Afterwards, the buds were immersed and agitated in 20% (w/v) of Clorox (5.25% w/v of sodium hypochlorite) added with drops of Tween 20 with constant agitation. Next, they washed with sterile distilled water for five times. Under aseptic conditions, bud scales peeled off and later trimmed to about 0.5 cm long. Medium complemented by various hormones used according to treatments in culture medium for the regeneration of shoots and create of roots from shoots multiplied for product seedlings.



Fig. 1. Location of Al-Mahwit governorate (Red point) (located in northwest of capital city of Yemen (Sana'a). It is located at an elevation of about 2000 meters above the sea level. The climate of Al-Mawhit is considered as *Aw* (tropical savanna climate with dry-winter characteristics) according to the Köppen-Geiger climate classification. The average temperature in Al Mahwit is 23.1 °C. The rainfall is around 1189 mm per year. (<https://en.climate-data.org/asia/yemen/al-mahwit-governorate/al-mahwit-54923/>).

Experimental design to induce multiple shoots

Two experiments were performed in the tissue culture laboratory of the University of Sana'a:

The first experiment was done to find the most powerful pH medium. To do so, an experiment based on completely randomized block design was conducted with four replicates. The MS (Murashige and Skoog 1962) medium was used + 1 mg L⁻¹ IBA at various pH levels (5.6, 5.8, 6.0). The right-most pH selected from this experiment was used in the second experiment.

In the second experiment, to test the best cytokinin type and level, a factorial experiment based on completely randomized block design was conducted with four replicates, including an experimental unit containing 10 test tubes. Two factors involved: i) Types of cytokinin: 6-benzylaminopurine (BAP) and Kiniten (Kin) (N6-furfuryladenine) and ii) Cytokinin concentrations (0, 1, 2 and 3 mg L⁻¹). The MS basal medium supplemented with above types and concentration of cytokinin with 1.0 mg L⁻¹ IBA and 3% sucrose, the medium solidified with

0.7% agar at pH 5.8 and sterilized by using autoclaving for 8 min.

Culture condition

Properly sterilized sprouts were longitudinally cutting into 5-10 mm in length in the laminar airflow cabinet, and directly cultured on shoot regeneration medium containing 15 ml of MS medium supplemented with various concentrations of BAP and Kin as per treatments. One piece inoculated in each tube containing sterile culture medium with different concentrations of plant growth regulators for the shoot and root induction and then transferred to growth room and allowed to grow in a controlled environment. The temperature of the growth room maintained at 25±2 °C with an air conditioner, light period of 16 h maintained with an intensity of 2000 lux for growth and cultivation development.

Measurements

After 20 days of first experiment (pH levels), and after 6 weeks of second experiment (cytokinin

types and levels), the shoot length, shoot number and root number was recorded for all culture tube. Rooted shoots removed from the culture tube and the root washed under running water to remove the agar. For hardening, the seedlings transferred to plastic pots containing peat moss and covered with polyethylene (Fig. 5A and B).

Data analysis

The data was statistically analyzed based on a RCBD, pH experiment was analysis as one way ANOVA with four replication, and the cytokinin experiment was analysis as a two-way ANOVA with four replicates using *Genstat 12* software. The least significant difference (LSD) was used for the comparison of means amongst the treatments.

Results

Effect of pH levels

The effect of pH levels on the average growth of the shoot and roots is show in Table 1. Medium with pH at 5.8 produced significantly more shoot and root than the other pH levels. It was the highest with about 152% and 141% on shoots length, 267% and 308% on shoots number and 276% and 159% on the root number as compared to their characteristics in the lowest 5.6 and highest 6 pH levels respectively. Meanwhile, the lowest values labelled at 5.6 pH level (Table 1, Fig. 2). The explant values were green without verification under all pH levels.

Table 1: Effect of different medium pH levels on shoot and root development of Yemeni local ginger variety from shoot-tip explants

pH Levels	Shoot length(cm)	No. Shoot	No. Root
5.6	5.0 ^b	1.5 ^b	6.5 ^c
5.8	7.6 ^a	4.0 ^a	18.0 ^a
6.0	5.4 ^b	1.3 ^b	11.3 ^b
LSD _{0.05}	1.287	0.8619	1.389

Means from four replicates for different medium pH levels (5.6, 5.8 and, 6.0) are presented. Different letters in the same column indicate significant differences ($p < 0.05$, Fisher's least significant difference test). All medium with different medium pH levels contain 1 mg L⁻¹ IBA.

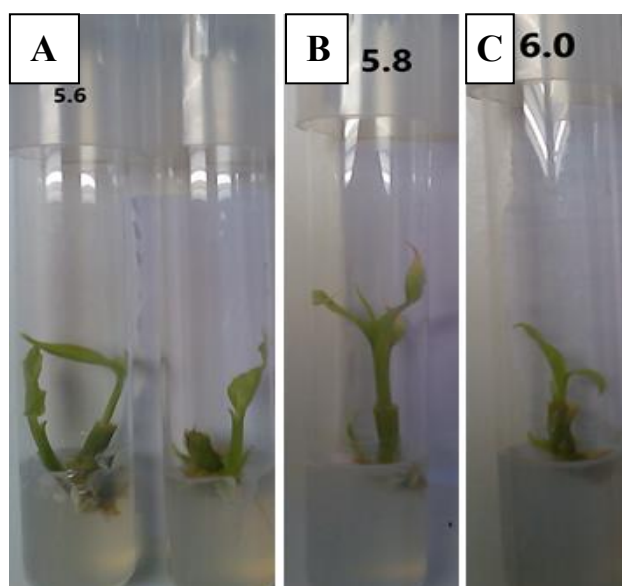


Fig 2: Effect of Different Levels of pH on Regeneration of Yemeni local ginger variety after 20 Days of culture. From the left: (A) : 5.6 , (B): 5.8 and (C) : 6.0

Effect of cytokinin type and concentration

In this study, cytokinin significantly affected the regeneration of local Yemeni ginger (Table. 2) and (Fig. 3 and 4). Overall, after 6 weeks of

culture, Kin caused significantly the highest shoot number, shoot length, and root number by about 168.9%, 116.6%, and 173.6%, respectively, as contrasted with BAP.

Table 2. Effect of Cytokinin types (BAP and Kin) and its concentrations on the numbers of shoots and roots of Yemeni local ginger variety after 6 weeks of culture

**Treatments		No. Shoot	Shoot height (cm)	No. Root
Cytokinin (H)				
	BAP	1.875 ^b	4.75 ^b	9.12 ^b
	Kin	3.167 ^a	5.54 ^a	15.83 ^a
LSD_{0.05}		0.3559	0.624	0.937
Concentration (C) mg L⁻¹				
	0*	1.33 ^c	3.00 ^a	7.00 ^a
	1	3.25 ^a	6.50 ^c	14.42 ^b
	2	3.33 ^a	6.25 ^c	14.83 ^b
	3	2.17 ^b	4.83 ^b	13.67 ^b
LSD_{0.05}		0.5033	0.882	1.325
H*C				
	BAP	0	1.33 ^c	3.00 ^c
	BAP	1	3.50 ^b	6.33 ^a
	BAP	2	1.33 ^c	5.00 ^b
	BAP	3	1.33 ^c	4.67 ^b
	Kin	0	1.33 ^c	3.00 ^c
	Kin	1	3.00 ^b	6.67 ^a
	Kin	2	5.33^a	7.50^a
	Kin	3	3.00 ^b	5.00 ^b
LSD_{0.05}		0.7117	1.247	1.873

*Control (without cytokinin). All medium contain the cytokinin treatments +1 mg L⁻¹ IBA, and pH is equal 5.8. Any two means in the same column for the same factor (Cytokinin types (H), Concentrations (C) or H*C) not followed by the same letter are significantly different ($p < 0.05$) using the analysis of variance and standard Fisher's protected LSD.

The data presented in Table 2 show the highest result of all parameters on MS medium supplemented with BAP or Kin at concentration of 2 mg L⁻¹

¹. The higher or lower concentrations than 2 mg L⁻¹ gave less significant results in all parameters with a similar concentration of 1 mg L⁻¹.

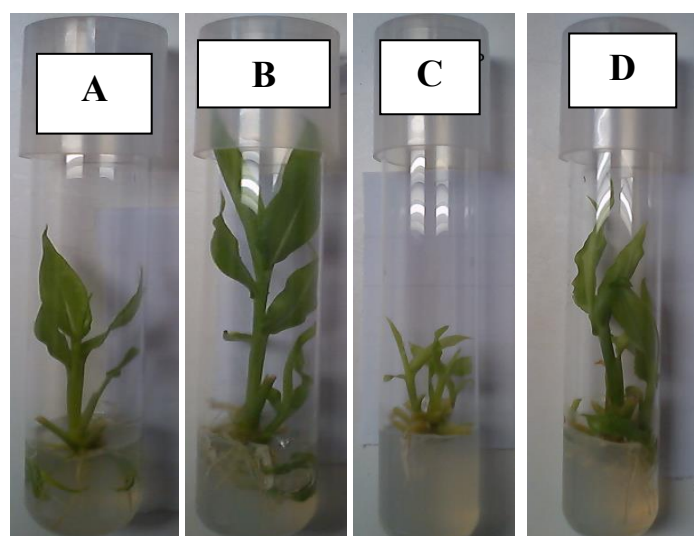


Fig. 3. *In vitro* micro-propagation of Yemeni local ginger variety, vegetative bud culture on MS fortified with different concentrations of BAP and multiple shooting and root induction after 6 weeks of culture. From the left (A) : 0.0 , (B): 1 mg L⁻¹, (C); 2 mg L⁻¹, (D): 3 mg L⁻¹

The significant difference between BAP and Kin is also clear in Table 2. One mg L⁻¹ of BAP or Kin provides the same influence with no significant difference against shoots number and shoots height. Nevertheless, the number of the root was higher in one mg L⁻¹ of Kin by about 43.7% than one mg L⁻¹ BAP (Table 2). BAP at 1.0 mg L⁻¹

proceeded in the highest multiple shoot length, shoot number and root number with about 210%, 262%, 168%, respectively, as compared with control treatment (with no hormones). There were no significant differences in on shoot number and shoot height at $p < 0.05$ between 1 and 2 mg/l of BAP.

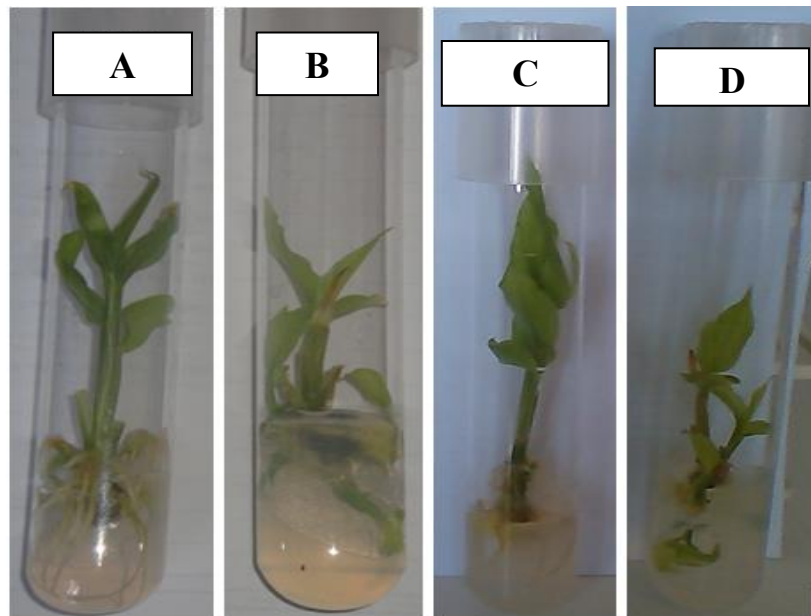


Fig. 4. *In vitro* micro-propagation of Yemeni local ginger variety, vegetative bud culture on MS fortified with different concentrations of Kin and multiple shooting and root induction after 6 weeks of culture. From the left: (A) : 0.0 mg L⁻¹, (B): 1 mg L⁻¹, (C); 2 mg L⁻¹ and (D): 3 mg L⁻¹

The impact of Kin on the formation of the shoots and roots was clear, the concentration of 2.0 mg L⁻¹ Kin produced the highest shoot number (5.3),

shoot height (7.5) and root number (22.8) by about 400%, 250% and 319% higher compared to the control treatment, respectively.

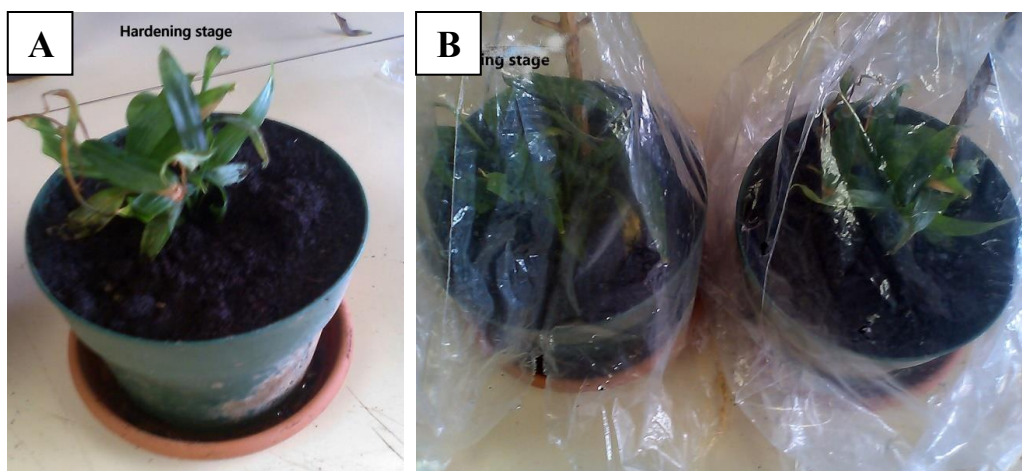


Fig. 5. Regenerated plant of Yemeni local ginger variety from sprout in plastic pot contain peat moss medium (A) and covered with polythene bag kept in lab for hardening (B).

Discussion

Micropropagation of plants *in vitro* mainly depends upon various factors and culture conditions. The medium pH is also a critical factor among them that correlated with plant growth under *in vitro* conditions. The optimization of culture media pH is a necessary but critical step to achieve rapid and efficient growth of plantlets by using *in vitro* practices (Sharma et al., 2018). In the present study, it was confirmed that the pH values play an important role in the manipulation or management of the induction result of ginger explants. This finding is consistent with previous research candidates by Rout and Das (1997) and Abdul Aziz et al. (2012).

Owen et al. (1991) stated that the pH of the media in tissue culture system could influence *in vitro* shoot multiplication, floral and secondary metabolites development, organogenesis, production of adventitious roots, and cell division. Whatever the case, based on the previous studies, pH media was installed 5.8 as we consider it as the optimum level in the current experiment like studies by Balachandran et al. (1990), Abbas et al. (2011), and Zahid et al. (2021).

As a matter of fact, plant growth regulators are one of the main factors affecting growth and development of explant in tissue culture. Cytokinins play major roles in shooting of different plants, either on foliage development *in vitro* (Rangan et al., 2004). Cytokinin stimulates cell division and increases the overall number of vegetative cells. It increases growth rate, stimulates the rate of metabolism of cells in the shoot, promotes storage of nutrients, increase active transport of nutrients and water, increase the flow of sugar and oxygen toward the site of syntheses (Rangan et al., 2004; Emami et al., 2011; Al-madhagi 2012).

The positive effect of MS medium augmented with 2 mg L⁻¹ Kin on increasing of shoots number, shoot height and root number was taken same direction with that found by Sharma and Singh (1997), who found the maximum number of shoots per bud during culture initiation in the medium containing 2 mg L⁻¹ Kin. Moreover, it is in accordance with Arildo et al. (2003) where Kin induces more shoots by explant of *Tabernaemontana fuchsiaeolia* L than the BAP. Also in agreement with Abu-Romman et al. (2015) who showed that Kin has most effective in inducing bud emergence from nodal explants of cucumber. Moreover, Hajare et al. (2021) found the numbers of multiple shoots was differing between potato cultivars where, in one cultivar was the highest in the MS medium containing 2.5 mg L⁻¹ Kin and other was when used PAB. Moreover, Buah et al. (2010) suggested that Kin has the capacity to induce more banana shoots

with increased concentrations at 7.5 mg L⁻¹. Hashemidehkordi et al. (2021) found that MS medium supplemented with 0.5 mg L⁻¹ (IAA) + 0.1 mg L⁻¹ Kin gave the highest rooting percentage and root number per explant of *Zantedeschia spp* and the highest root length was observed in 1.0 mg L⁻¹ IAA + 0.1 mg L⁻¹ Kin.

In contrast to our results, BAP was reported in many studies such as by Balachandran et al. (1990), Abbas et al. (2011) and Zahid et al. (2021) as the most effective cytokinin for shoot induction and multiplication in ginger. As well in other plant species such as *Daphne mezereum* L in the study by Nowakowska et al. (2019), who found that the highest number of shoots produced on MS medium with 1 mg L⁻¹ BA and 0.1 mg L⁻¹ NAA than the other types of cytokinins (meta-Topolin and zeatin).

The differences found in the relative strengths of the different cytokinin types in inducing shoots in ginger are probably due to a mixture of factors such as stability, mobility, the rate of conjugation and oxidation of the hormones (Buah et al., 2010). This is also due to the antagonistic interaction between cytokinin and auxin in the MS medium (Kurepa et al., 2019). Where, the auxin/cytokinin ratio is the major physiological factor affecting shoot/root initiation (Rangan et al., 2004; Amin et al., 2007). It is clear that the cytokinin types have also major roles in this matter. Kurepa et al. (2019) concluded that *in vitro* auxin inhibits the response to low concentrations of cytokinin as a consequence of the BA, but is ineffective if the cytokinin content is high. The application of exogenous cytokinins *in vitro* resulted in an increase in the cytokinin content of the shoots, and application of zeatin and dihydrozeatin also detected in the newly-formed shoots. Application of BA to the media led to a transition from isoprenoid cytokinins to aromatic cytokinins in the shoots (Quiala et al., 2014).

This may clearly argue that the role of auxin on rooting, and increasing or decreasing endogenous cytokinin may inhibit the role of auxin. Auxin and cytokinin play an important regulatory role in the plant, not only in apical dominance but also in many other correlated events as well and cytokinins exudates in the intact plants are under the control of the polar auxin transport system (Bangerth, 1994). Supporting to this role, Nirmal (1997) found that addition of NAA alone gives the highest root induction, whereas BAP alone at higher concentration (4 mg L⁻¹) induces only multiple shoots and rarely roots. Kambaska and Santilata (2009) found that the NAA is more effective than IBA in induction of ginger rooting as days required for rooting is lower by half of days as responsive to IBA. Adding 0.5 mg L⁻¹ NAA

with BAP (2 mg L⁻¹) improve response over BAP alone and also it has been suggested that the combinations of BAP and NAA are needed for producing more number of multiple shoots on *Zingiber officinale* (Hoque et al., 1977 ; Hashim et al., 1998 ; Noguchi and Yamakawa., 1998).

In our experiment, the highest level of BAP and Kin (3 mg L⁻¹) gives the lowest result in all parameters with a similar concentration of 1 mg L⁻¹. These results are consistent with findings from Inden et al. (1988); Sharma and Singh (1997) and Anjumanara et al. (2003), who reported that high concentrations of cytokinin reduced shoot elongation, reduced rooting and caused genetic instability in micro-propagated ginger plants. Furthermore, Sharma and Singh (1997) indicated that low concentrations of Kin have a positive effect on *in vitro* shoot multiplication, where, plantlets obtained at lower concentrations of Kin had well-developed shoots, opened leaves and sparse and thin rooting, and at higher concentrations of Kin, less shoot elongation, folded leaves and very rare rooting were observed. This result also interpreted before by Teisson and Cote (1985), when over exposure to high level of cytokinins may lead to vitrification. In fact, the function of cytokinins or auxins is perspicuous on the

References

- Abbas Mohamed S, Taha Hussein S, Aly Usama I, El-Shabrawi Hattem M, and Gaber, El-Sayed I. 2011. *In vitro* propagation of ginger (*Zingiber officinale* Rosco). Journal of Genetic Engineering and Biotechnology 9, (2) 165-172. <https://doi.org/10.1016/j.jgeb.2011.11.002>
- Abdul Aziz, Monzur Hossain, Rafiul Islam. 2012. *In Vitro* Organogenesis in Ginger: *In vitro* Organogenesis and Plant Regeneration of Ginger (*Zingiber officinale* Rosc.) . Lap Lambert Academic Publishing. ISBN 13: 9783848404957 Paperback - March 14, 2012
- Abu-Romman, Saeid M, Khaldoun A, Al-Hadid Abdullah, Arabiyyat R. 2015. Kinetin Is the Most Effective Cytokinin on Shoot Multiplication from Cucumber. Journal of Agricultural Science 7(10), 159-165. <https://doi.org/10.5539/JAS.V7N10P159>
- Al-Madhagi Isam. 2012. Influence of Photoperiod and Exogenous Hormone on Growth and Development of Strawberry (*Fragaria X Ananassa* Duch.) PhD Thesis, Universiti University Malaysia Terengganu, Malaysia.
- Amin A A , Rashad M E S , EL-Abagy H M H. 2007. Physiological effect of Indole - 3-butyric acid and salicylic acid on growth, Yield and Chemical constituents of onion plants. Journal of Applied Sciences Research 3(11): 1554-1563.
- Anjumanara Khatun, Shamima Nasrin, Tojammal M Hossain. 2003. Large Scale Multiplication of Ginger (*Zingiber Officinale* Rosc.) From Shoot-tip Culture. Journal of Biological Sciences 3, 59-64.

concentration of other endogenous hormones in plants or in exogenous application (Rangan et al., 2004; Al-madhagi 2012).

Conclusion

It could be concluded that Kinetin is the best hormone for propagation of local Yemeni gainer with the concentration of 2 mg L⁻¹. Even though the role of cytokinin has been widely understood, the difference in the role of cytokinin types remains uncertain. It can be suggested that the effects of different types and concentrations of hormones should be analyzed on the level of endogenous hormone in tissue culture.

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Conflict of Interests

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<https://doi.org/10.3923/jbs.2003.59.64>

Arildo José Braz de Oliveira, Vanda Marilza de Carvalho, Alexandre Ferreira, Fernando Y Sato, Maria de Fátima Pires da Silva Machado. 2003. *In vitro* multiplication of *Tabernaemontana fuchsiaeifolia* L. Revista Arvore, 27 (4): 421-425. <https://doi.org/10.1590/S0100-67622003000400001>

Ayenew Biruk, Wondyifraw Tefera, B. Kassahun. 2012. *In vitro* propagation of Ethiopian ginger (*Zingiber officinale* Rosc.) cultivars: Evaluation of explant types and hormone combinations. African Journal of Biotechnology 11(16), 3911-3918 <https://doi.org/10.5897/AJB10.1962>

Balachandran S M , Bhat S R, Chandel K P S. 1990. *In vitro* clonal multiplication of turmeric (*Curcuma sp.*) and ginger (*Zingiber officinale* Rosc.). Plant Cell Reports, 8, 521-524. <https://doi.org/10.1007/BF00820200>

Bangerth F. (1994). Response of cytokinin concentration in the xylem exudate of bean (*Phaseolus vulgaris* L.) plants to decapitation and auxin treatment, and relationship to apical dominance. Planta 194(3), 439-442. <https://doi.org/10.1007/BF00197546>

Behera K K, Santilata Sahoo . 2009. An efficient method of micropropagation of ginger (*Zingiber officinale* Rosc. cv. Suprava and Suruchi) through *in vitro* rhizome bud culture. Indian Journal of Plant Physiology 14 (2) , 162-168

- Bhattacharyya P, Kumaria S, Diengdoh R, Tandon P. 2014. Genetic stability and phytochemical analysis of the *in vitro* regenerated plants of *dendrobium nobile* (lindl) an endangered medicinal orchid. *Meta Gene* 2, 489-504. <https://doi.org/10.1016/j.mgene.2014.06.003>
- Buah J N, Danso E, Taah K J, Abole E A, Bediako E A, Asiedu J, Baidoo R . 2010. The Effects of Different Concentrations Cytokinins on the *in vitro* Multiplication of Plantain (*Musa* sp.). *Biotechnology* 9, 343-347. <https://doi.org//10.3923/biotech.2010.343.347>
- Charlwood K A, Brown S, Charlwood B V. 1988. The accumulation of flavour compounds by cultivars of *Zingiber officinale*. Eds Richard J R, Michael J C, Rhodes Manipulating Secondary Metabolites in Culture. AFRC Institute of Food Research, Norwich, U.K, 195–200.
- Quiala E., Marco V, Jiménez-Tello, Raúl Barbón , Maité Chávez , Manuel de Feria , Mariana La O, Marta Pérez. 2014 . Influence of 6-Benzyladenine and gelling agent on the reduction of hyperhydricity in *Tectona grandis* L. *Revista Colombiana de Biotecnología* 16 , 129-136. <https://doi.org/10.15446/rev.colomb.biote.v16n1.44279>
- Emami H , Saeidnia M , Hatamzadeh A , Bakhshi D, Ghorbani E. 2011. The Effect of Gibberellic Acid and Benzyladenine in Growth and Flowering of Lily (*Lilium longiflorum*). *Advances in Environmental Biology* 5(7), 1606-1611.
- Eshghi Khas Mostafa, Abbasifar Ahmadreza, Valizadeh Kaji Babak. Eshghi Khas, M., Abbasifar, A. & ValizadehKaji, B. (2020). Optimization of *in vitro* Propagation of Purple Passion Fruit (*Passiflora edulis*), an Important Medicinal and Ornamental Plant. *International Journal of Horticultural Science and Technology* 7(3):), 305-314.
- FAOSTAT Agricultural Statistics Database <http://www.fao.org>. 2013
- Hajare S T, Chauhan N M, Kassa G. 2021. Effect of Growth Regulators on *in Vitro* Micropropagation of Potato (*Solanum tuberosum* L.) Gudiene and Belete Varieties from Ethiopia. *The Scientific World Journal* Volume 2021, Article ID 5928769, 8 pages. <https://doi.org/10.1155/2021/5928769>
- Hashemidehkordi Elaheh , Seyed N M , Pejman A. 2021. An Efficient *in vitro* Propagation Protocol Of Pot Calla Lily (*Zantedeschia* spp *cv.* Orania and sunclub) via tuber production. *International Journal of Horticultural Science and Technology* 8,(4) 343-351. <http://dx.doi.org/10.22059/ijhst.2021.317458.436>
- Hossain A , Hassan L, Patwary A, Mia M, Ahmad S D, Shah A, Batool F. 2010. Establishment of a suitable and reproducible protocol for *in vitro* regeneration of ginger (*Zingiber Officinale* Roscoe). *Pakistan Journal of Botany* 42: 1065-1074.
- Inden H, Asahira T, Hirano A. 1988 . Micropropagation of Ginger. *Acta Horticulturae* 230, 177-184. <https://doi.org/10.17660/ActaHortic.1988.230.20>
- Kishore N , Dwivedi . 1992. Zerumbone: a potential fungi toxic agent isolated from *Zingiber cassumunar* Roxb. *Micopathologia*, 120(3), 155-159.
- Kurepa J, Shull TE, Smalle JA. 2019. Antagonistic activity of auxin and cytokinin in shoot and root organs. *Plant Direct* 3 , 1-9. <https://doi.org/10.1002/pld3.121>
- Naderi Boldaji H , Dianati Daylami S , Aliniaiefard S , Norouzi M. 2021. Efficient Method for Direct Embryogenesis in Phalaenopsis Orchid. *International Journal of Horticultural Science and Technology* 8(1), 37-50. <https://dx.doi.org/10.22059/ijhst.2020.296696.339>
- Nair K P. 2019. Turmeric (*Curcuma longa* L.) and Ginger (*Zingiber officinale* Rosc.)-World's Invaluable Medicinal Spices: The Agronomy and Economy of Turmeric and Ginger; Springer Nature: Basel, Switzerland ISBN 9783030291884
- Nirmal Babu K. 1997. *In vitro* studies in ginger, *Zingiber officinale* Rosc. Unpublished Ph.D. Thesis, University of Calicut, Kerala, India.
- Nowakowska K, Pacholczak A, Tepper W. 2019. The effect of selected growth regulators and culture media on regeneration of daphne mezereum l.'alba'. *Rendiconti Lincei. Scienze Fisiche e Naturali* 30(1), 197-205. <https://doi.org/10.1007/s12210-019-00777-w>
- Olivier J J. 1996 . The initiation and multiplication of ginger (*Zingiber officinale* Rosc.) in tissue culture. In *lightings bulletin* -Instituut VIN Tropiese Subtropiese Gewasse, 291, 10-11.
- Owen H R, Wengerd D, Miller A R. 1991. Culture medium pH is influenced by basal medium, carbohydrate source, gelling agent, activated charcoal, and medium storage method. *Plant cell reports* 10(11), 583–586. <https://doi.org/10.1007/BF00232516>
- Ozaki Y, Kawahara N, and Harada M. 1991. Anti-inflammatory effect of *Zingiber cassumunar* Roxb. and its active principles. *Chemical and pharmaceutical bulletin* Chem. Pharm. Bull. 39(9), 2353- 2356. <https://doi.org/10.1248/cpb.39.2353>
- Rangan R, Purohit S S, Prasad V. 2004. *Plant hormones Action and Application*. India, Agrobios. ISBN 13: 9788177541649
- Rout G R, Das P. 1997. *In vitro* Organogenesis in Ginger (*Zingiber officinale* Rosc.)" *Journal of Herbs, Spices & Medicinal Plants* 4(4) , 41-51. https://doi.org/10.1300/J044v04n04_05
- Sakamura F, Suga T. 1989. *Zingiber officinale* Roscoe (Ginger): *In Vitro* Propagation and the Production of Volatile Constituents. In: Bajaj Y.P.S. (eds) *Medicinal and Aromatic Plants II. Biotechnology in Agriculture and Forestry*, vol 7. Springer, Berlin, Heidelberg. https://doi.org/10.1007/978-3-642-73617-9_29
- Sakamura F, Ogihara K , Suga T , Taniguchi K , Tanaka R. 1986. Volatile constituents of *Zingiber officinale*

rhizome produced by *in vitro* shoot tip culture. *Phytochemistry* 25 (6), 1333-1335 . [https://doi.org/10.1016/S0031-9422\(00\)81284-4](https://doi.org/10.1016/S0031-9422(00)81284-4)

Sharma M K , Chaudhary R, Kureel R, Sengar R. 2018. Effects of culture media pH on *In Vitro* shoot multiplication in sugarcane. *International Journal of Chemical Studies* 6(2), 1308-1310

Sharma T R, Singh B M. 1997. High-frequency *in vitro* multiplication of disease-free *Zingiber officinale* Rosc. *Plant Cell Reports* 17(1), 68-72. <https://doi.org/10.1007/s002990050354>

Teisson C, Cote F X . 1985. High-tech and Micropropagation. In: *Biotechnology in Agriculture and Forestry*, Bajaj, Y.P.S. . (Ed.). Bajaj Y P S. Springer Verlag, Berlin, Heidelberg, ISBN: 3-540-61606-3, pp: 254.

Zahid N A , Jaafar H Z E , Hakiman M. 2021. Micropropagation of Ginger (*Zingiber officinale* Roscoe) 'Bentong' and Evaluation of Its Secondary Metabolites and Antioxidant Activities Compared with the Conventionally Propagated Plant. *Plants* 10, 630. <https://doi.org/10.3390/plants10040630>

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